

# An overview of General Anesthesia in Laboratory Animals

## Falah Mahmood Hameed, Ali Wasfi Sadeq

College of Veterinary Medicine, University of Kerbala, Kerbala, Iraq. Corresponding author: falah.m@uokerbala.edu.iq, ali.w@uokerbala.edu.iq

Received: 8/5/2025 Accepted: 25/5/2025 Published: 3/7/2025

Abstract— This review article examines general anesthesia in laboratory animals. Administering anesthesia in any species necessitates careful planning, execution, and resources to guarantee a humane and successful result. Anesthesia is crucial elements in lab animal, and they must be included as fundamental parts of all lab animal programs. It stops avoidable suffering caused by different experimental methods (1). Unmanaged or neglected pain, leading to the release of uncontrolled substances. Ultimately, it may result in a range of undesirable alterations in its body. In the end, this will impact the results of the experiment. Due to these factors, the responsible application of anaesthesia and analgesia is a scientific and ethical necessity (2). Anesthesia refers to a condition of unconsciousness, and its elements include analgesia (relief from pain), amnesia (memory loss), and immobilization. The medication employed for anaesthesia typically produces different effects in each of these regions. Certain medications might be utilized on their own to obtain all three outcomes. Some possess solely analgesic or sedative effects (3).

Keywords: General anaesthetic agents, anaesthetic restraint, lab animal

#### INTRODUCTION

addition to uses in regenerative medicine and surgery, animals of Laboratory are used in clinical experimental studies, (medication pre-market testing or medicalsurgical gadget assessment). The anaesthesia techniques may significantly change the outcomes of experiments and have an impact on the survival rates of lab animals. There isn't yet a commonly accepted anaesthetic regimen for any particular type of laboratory animal. The most often utilised model in a variety of research domains, including imaging, regenerative medicine, and organ transplantation, is the murine species, which includes rats and mice (4). Because animals typically refuse to participate during operations like therapy or illness assessment, anaesthesia is commonly used on them. It is just the absence of an animal's response to unpleasant stimuli, regardless of whether a consciousness loss occurs (1). A major purpose of anaesthesia is to keep animals' physiological processes as near to normal as possible while immobilising it and preventing it

from sensing unpleasant stimuli. An effective aesthetic medication lowers bronchial and salivary secretions, eases preoperative discomfort in animals, and ensures a smooth induction and recovery. According to (5), an aesthetic impact that lasts anywhere from thirty minutes to an hour is produced by an anaesthetic drug that provides a short duration of anaesthesia. Three essential classic elements of anaesthesia are narcosis, muscular relaxation, and pain relief. Before undergoing traumatic treatments like surgery, retro-orbital haemorrhage, or heart punctures, laboratory animals must be put to sleep. The animals will be far less stressed and in discomfort if the researcher is knowledgeable and competent in anaesthesia. Anaesthetics may be administered by injection or inhalation (6).

## 1.1. Goals of Anaesthesia:

- To carry out a surgical procedure for research aims
- Gain physiological stability for the processes
- Restore normal physiological function.
- To avoid discomfort and suffering during the process
- To ensure stability (maintain the catheter etc.)
- To ensure optimal conditions for surgical (Relaxation of muscle)

# 1.2 The ideal anaesthetic/analgesic regimen has to fulfil a number of requirements:

- 1. Lessen any pain or suffering brought on by handling or giving
- 2. Be precisely titratable to guarantee that animals get enough anaesthesia to render them unconscious and immobile and to stop them from feeling pain without causing haemodynamic instability.
- 3. To prevent the sensitisation of pain pathways and lower the overall dose of general anaesthetic required for the treatment, provide preemptive analgesia to make sure that pain



# "Kerbala Journal of Veterinary Medical Sciences, Vol. 1, Supplement I, 2025" "Proceedings of the 6th Scientific Conference, College of Veterinary Medicine, University of Karbala"

management starts while the general anaesthetic is still taking effect.

- 4. Do not disrupt the research objectives.
- 5. Avoid causing unwanted intra- or post-operative complications.
- 6. Be suitable for existing equipment, other drugs, and staff education.
- 1.3. Pre-anaesthetic Both the anaesthetist and the animal benefit from pre-anaesthesia or pre-medication, which makes it easier to induce and maintain anaesthesia and increases the animal's safety and comfort. In order to reduce anxiety and prepare for general anaesthesia, pre-medication is often administered before to invasive operations like surgery (Anaesthesia Protocols in Lab Animal Scientific Use (7).

#### 1.4 Sedation

Animals under sedative have calm and relaxing impact that lowers their activity levels and decreases their sensitivity to their environment. When administered as a premedication before to the onset of anaesthesia, it lowers body temperature and reduces the quantity of injectable anaesthesia by 20% to 50%. Premedication's primary goal is to reduce patients' anxiety and prepare them for general of anaesthesia (8).

## 1.5 Types of Pre-anesthetic medication

## 1.5.1 Agents with Anticholinergic Properties

Anticholinergics have long been used as premedication to reduce bronchial and salivary secretions. These medications, which include glycopyrrolate and atropine sulphate, may inhibit parasympathetic impulses. In some cases, it is essential to decrease bronchial secretions in order to avoid airway blockage while sedating animals (8).

## 1.5.2 Sedative Anaesthesia

This results in a deeper level of sedation, which intensifies sleepiness and inhibits both conditional (emotions, motives) and unconditional (automatic instincts) responses. Barbiturates (phenobarbital, barbital), Alpha 2 agonists (xylazine, detomidine), benzodiazepines (diazepam), and chloral hydrate are among the medications that have a shorter duration of effect (9).

## 1.5.3 Tranquilizing sedatives

Deeper sedation follows, which exacerbates drowsiness and suppresses both unconditional (automatic instincts) and conditional (emotions, motivations) reactions. Among the drugs with a shorter half-life include barbiturates (phenobarbital,

barbital), chloral hydrate, benzodiazepines (diazepam), and Alpha 2 agonists (xylazine, detomidine) (9).

## 1.6 Classification of anaesthetic drugs

Anaesthesia for laboratory animals can be generally categorized into:

- 1. Local A.
- 2. Regional A.
- 3. Sedatives
- 4. General Anaesthesia by inhalation /parenteral

## 1.7 Anaesthesia can be divided into four stages.

However, there is an incremental change from one phase to the next. From the start of anaesthesia until the recovery phase, it is crucial to regularly check the anaesthesia level. Making ensuring the animal is not too anaesthetised throughout the procedure is just as important as making sure it is sufficiently anaesthetised to avoid suffering. By monitoring reflex responses, Patterns of breathing and length, pulse, blood pressure, and responses to potentially harmful (painful) stimuli one may readily determine the degree of anaesthesia (10).

## 1.8 Toxic effect of anaesthesia

Aesthetic monitoring encompasses more than merely electrical monitoring equipment. Muscle movements or conscious reactions, especially Painful stimuli can be used to determine the level of anaesthesia. This can be determined by examining how an animal reacts to any painful event. When confronted with unpleasant stimuli such as pinching a toe or foot pad, discomfort can be expressed by raising the head and kicking a leg (11). Light anaesthesia in experimental animals causes the loss of the righting reflex (when a rat is flipped onto its back, it swiftly reorients itself) and immobility, yet the animal can still react strongly to potentially painful stimuli (12). The significance of anaesthetic monitoring devices Vital insights may be obtained from anaesthetic equipment, however for improved animal supervision, monitoring techniques should be tailored. Electrical anaesthetic devices are useful because they provide a better knowledge of anaesthesia; nevertheless, if the information is not used appropriately, they do not increase anaesthetic safety (1).

## 1.10 Examination of reflexes

The reflex arc in the spinal cord controls reflexes, which are automatic bodily reactions brought on by a stimulus. The attendance of reflexes cannot always indicate that having pain; certain reflexes may be absent while others may continue to function under anaesthesia. Reflexes are the sole indicators of an animal's level of anaesthesia if physiological parameters in response to painful stimuli cannot be tracked. According to



## "Kerbala Journal of Veterinary Medical Sciences, Vol. 1, Supplement I, 2025"

## "Proceedings of the 6th Scientific Conference, College of Veterinary Medicine, University of Karbala"

(13), It is possible that an animal with a positive response is aware of discomfort and is not fully anesthetized.

### 1.11 Four phases of anaesthesia

- 1. Induction phase: The creature stays awake and skills a slight degree of analgesia and sedative.
- 2. Excitation stage: The organism loses awareness and exhibits enhanced reflex reactions and mucus secretion.
- 3. Surgical stage: The animal's breathing becomes slow and deeply, and the eyelid and corneal responses vanish. The reflex response to surgical or other stimuli decreases.
- 4. Hypnotic or toxic stage: During this stage, critical brain areas become less active, resulting in decreased cardiac output and breathing, eventually leading to cessation. The pupils are completely dilated & fail to react to light. This stage can be noticed when animals are euthanized with large quantities of anaesthetic medicines.

#### 1.12 Anaesthesia method

- Intravenous
- Intra-muscular
- Intraperitoneal
- · Subcutáneo
- by mouth
- Inhaling

## 1.13 Restraint of animals physically

A variety of physical restraint techniques are necessary to initiate anaesthesia. There are restraint devices, but securing the animal in them takes longer than just holding it. It's crucial to keep in mind that, in order to reduce pain, physical restraint should be short (5). When it comes to anaesthesia, intravenous medication delivery accuracy is crucial. Techniques to reduce the animal's pain and suffering must be well-mastered by researchers. To do this, it is thus crucial to understand appropriate anaesthesia and intravenous procedures (6).

## CONCLUSION

Inadequate documentation of anaesthesia practices in experimental research was found by this systematic study. A small number of lab animals are always used in the investigation. This analysis also highlights the critical necessity for rules in experimental animal research; to guarantee the uniformity and calibre of animal research, certain anaesthesia and euthanasia methods should be taken into account and documented in subsequent studies. It is crucial to translate experimental findings into (future) therapeutic applications.

## **REFERENCES:**

- 1) 1. Flecknell, P. A. (1993). Review article anaesthesia of animals for biomedical research. British Journal of Anaesthesia, 71(6), 885–894.
- 2) 2. Hohlrieder M, Kaufmann M, Moritz M, Wenzel V. Management of accidental hypothermia. Anaesthesist. 2007;56:805–11. doi: 10.1007/s00101-007-1206-9.3. Whelan, G., & Flecknell, P. A. (1992). The assessment

- of depth of anaesthesia in animals and man. October 1991, 153–162.
- 3) 3. Olsen AK, Smith DF. Anaesthesia for positron emission tomography scanning of animal brains. Laboratory Animals. 2013;47:12–8. doi: 10.1258/la.2012.011173.
- **4**) 4. Swindle MM, Smith AC, Hepburn BJ. Swine as models in experimental surgery. J Invest Surg 1988; 1: 65-79.
- 5) 5. Cicero, L., Fazzotta, S., Palumbo, V. D., Cassata, G., Ignazio, A., & Monte, L. (2018). Anesthesia protocols in laboratory animals used for scientific purposes. 89(3), 337–342.
- 6) 6. Sotocinal SG, Sorge RE, Zaloum A, et al. The Rat Grimace Scale: a partially automated method for quantifying pain in the laboratory rat via facial expressions. Molecular Pain. 2011;7:55. doi: 10.1186/1744-8069-7-55.
- 7) 7. Hildebrandt IJ, Su H, Weber WA. Anaesthesia and other considerations for in vivo imaging of small animals. ILAR J. 2008;49:17–26.7.
- 8) 8. Ahmadi-Noorbakhsh, S., Farajli Abbasi, M., Ghasemi, M., Bayat, G., Davoodian, N., Sharif-Paghaleh, E., Poormoosavi, S. M., Rafizadeh, M., Maleki, M., Shirzad-Aski, H., Kargar Jahromi, H., Dadkhah, M., Khalvati, B., Safari, T., Behmanesh, M. A., Khoshnam, S. E., Houshmand, G., & Talaei, S. A. (2022). Anesthesia and analgesia for common research models of adult mice. Laboratory animal research, 38(1), 40. https://doi.org/10.1186/s42826-022-00150-3
- 9) 9. Gargiulo, S., Greco, A., Gramanzini, M., Esposito, S., Affuso, A., Brunetti, A., & Vesce, G. (2012). Mice anesthesia, analgesia, and care, Part I: anesthetic considerations in preclinical research. ILAR journal, 53(1), E55–E69. https://doi.org/10.1093/ilar.53.1.55
- 10) 10. Pehböck, D., Dietrich, H., Klima, G., Paal, P., Lindner, K. H., & Wenzel, V. (2015). Anesthesia in swine: optimizing a laboratory model to optimize translational research. Der Anaesthesist, 64(1), 65–70. https://doi.org/10.1007/s00101-014-2371-2
- 11) 11. Konno K, Itano N, Ogawa T, Hatakeyama M, Shioya K, Kasai N. New visible endotracheal intubation methodusing the endoscope system for mice inhalational anesthesia. J. Vet. Med. Sci. 2014;76(10):1375–81. doi: 10.1292/jvms.13-0647.
- 12) 12. Zhang, X., Sha, Y., Liu, X., Xun, M., Pang, J., He, W., ... Liu, J. (2025). The role of strial macrophages in the maintenance of the blood–labyrinth barrier. Academia Biology, 3(2). https://doi.org/10.20935/AcadBio1765
- 13) 13. Konno K, Shiotani Y, Itano N, et al. Visible, Safe and Certain Endotracheal Intubation Using Endoscope System and Inhalation Anesthesia for Rats. J Vet Med Sci. 2014;76(10):1375–81. doi: 10.1292/jvms.14-0146